



# **POINT-OF-CARE / DECENTRALIZED CAR-T CELL PRODUCTION**

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# CAR-T cells

## Belgian legislation

Human Bodily Material  
Collected by MD in hospitals

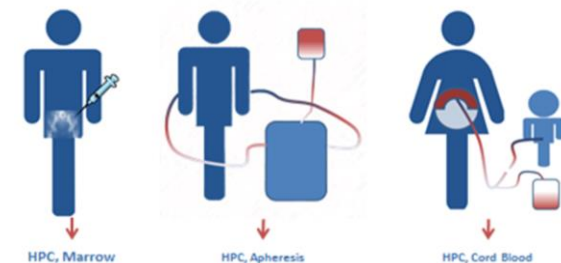
### Scientific research

Biobank  
No therapeutic use in humans

### Therapeutic use

Non-substantial manipulation

- Cell & tissue bank (hospital)
- Intermediate structure (commercial)

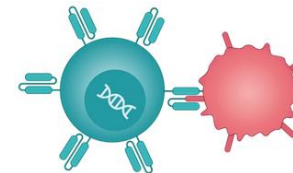
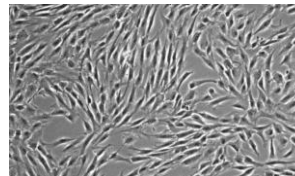


Substantial transformation

- ATMP : **A**dvanced **T**herapeutic **M**edicinal **P**roduct  
(cells, genes, tissues, combinations)
- GMP : **G**ood **M**anufacturing **P**ractice production



Loi du 19 décembre 2008 relative à l'obtention et à l'utilisation de matériel corporel humain destiné à des applications médicales humaines ou à des fins de recherche scientifique



# Liège Laboratory of Cell & Gene Therapy

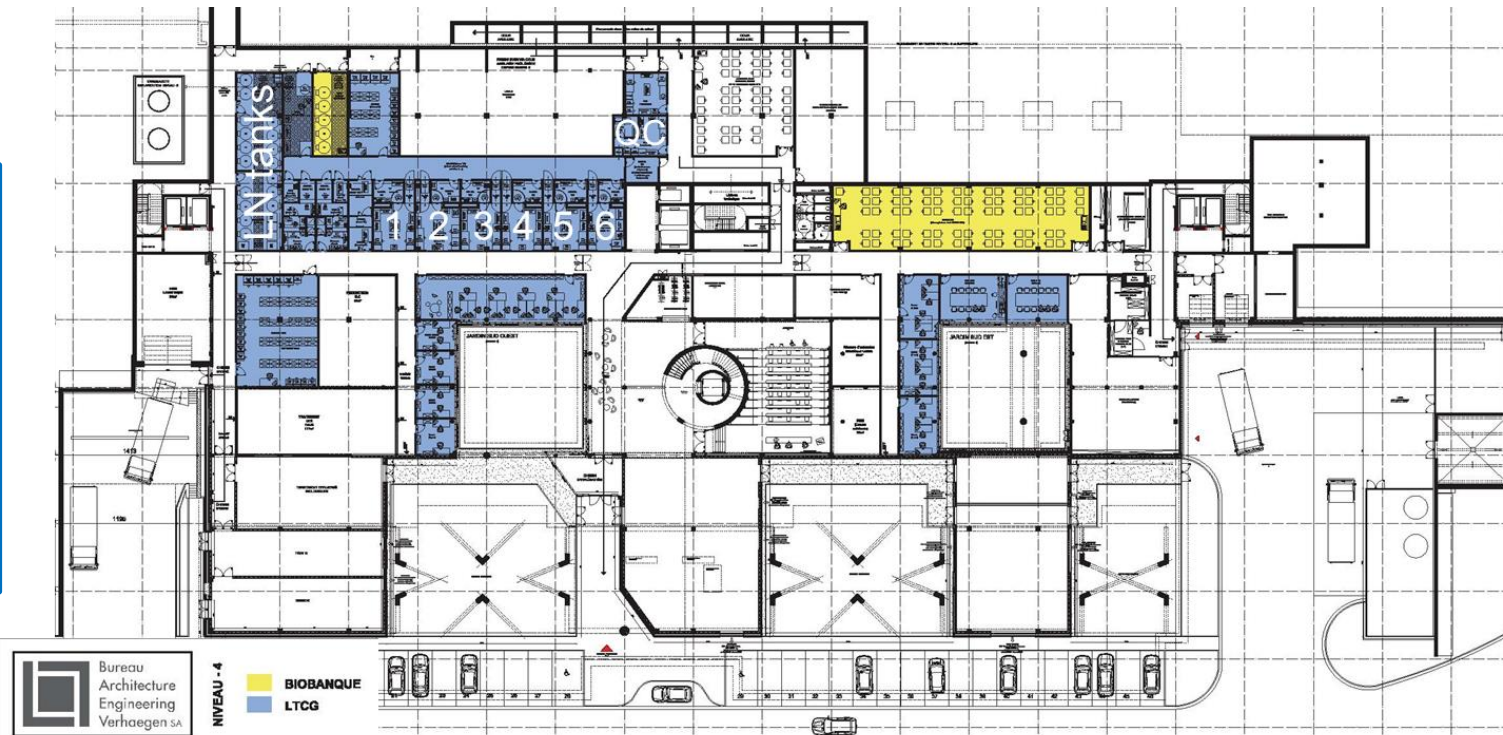
## Production & QC facilities



**ICAB**  
Institut de  
Cancérologie  
Arsène Burny


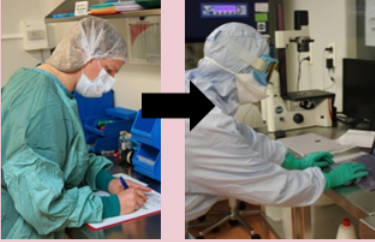
**LTCG**

- 6 GMP facilities
- 1 QC lab
- 75 m<sup>2</sup> of LN storage
- Storage areas
- Offices



# Liège Laboratory of Cell & Gene Therapy

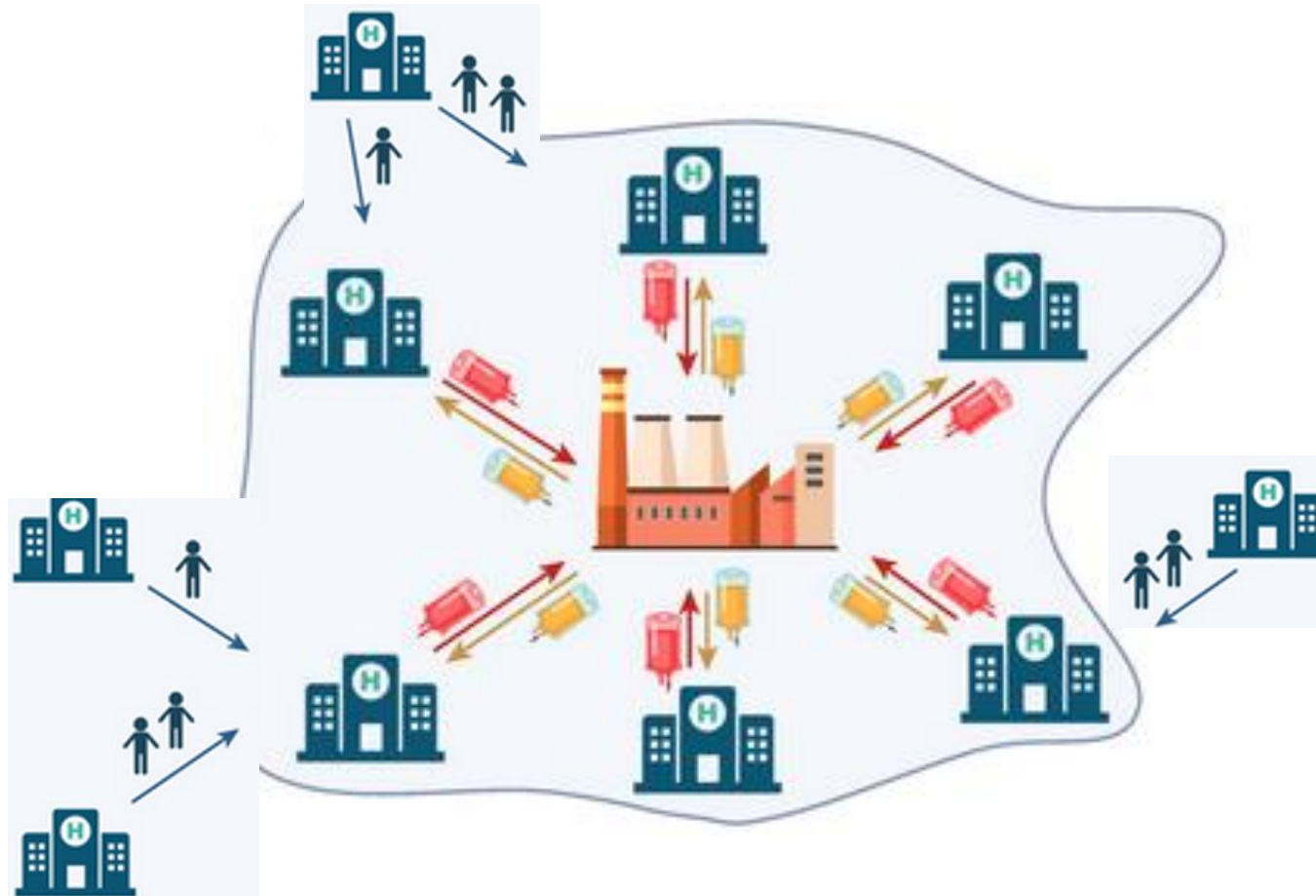
## MSC : from clinical grade to GMP

<p><b>Personnel</b></p> <p>+ </p> <ul style="list-style-type: none"> <li>• QC ≠ Production</li> <li>• Instrumentation</li> <li>• Personne qualifiée</li> </ul>	<p><b>Locaux</b></p> <p>Classe C ↓ Classe B</p>	<p><b>Habillage</b></p> 	<p><b>Monitoring environnemental</b></p> <p>↑↑↑↑</p> <p>Fréquence, Nombre et type de prélèvements</p>
<p><b>Documentation</b></p> <p>+  • Déviations • OOS • CCR GMP = Generate more paper</p>	<p><b>Média simulation</b></p> <p>+ <b>Média simulation</b></p> <ul style="list-style-type: none"> <li>• 3X nouveau process</li> <li>• 2X/an par process</li> <li>• 1X/an par opérateur</li> </ul>	<p><b>Validation</b></p> <p>Validation du process ↓ Validation process, contrôles de qualité, nettoyage Protocoles &amp; rapports</p>	<p><b>Réactifs &amp; Consommables</b></p> <p>+ Quarantaine/libération (Spécifications) Evaluation fournisseurs</p>
<p><b>Equipements</b></p> <p>Maintenance annuelle ↓ Procédures et IQ/OQ/PQ</p>	<p><b>Echantillons rétention et référence</b></p> <p>+ d'échantillons (Identification et analyses)</p>	<p><b>Contrôles de qualité</b></p> <p>Changement de sous-traitants (certifiés GMP)</p>	<p><b>Process</b></p> <ul style="list-style-type: none"> <li>•Retrait antibiotiques</li> <li>•Trypsine recombinante (porcine)</li> <li>•Cellstacks</li> </ul>



# CAR-T cells

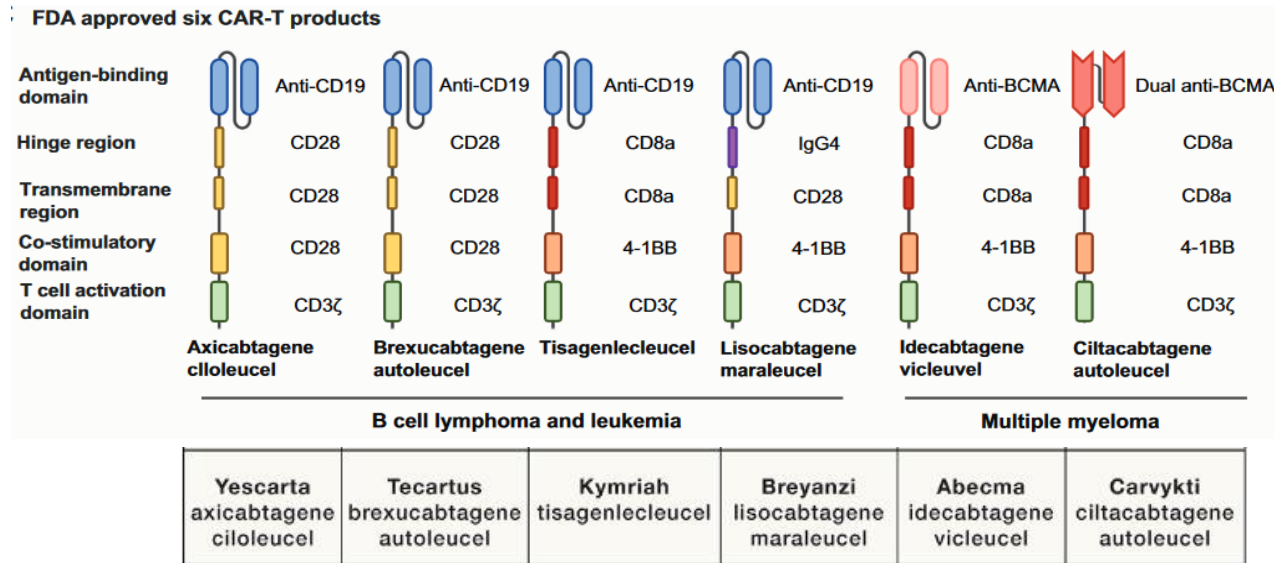
## Classical commercial CAR-T cell production



# CAR-T cells

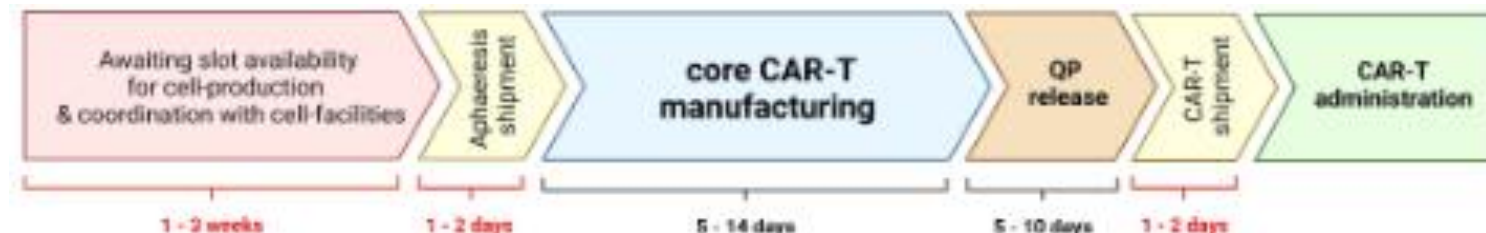
## B-cell malignancies : CAR-T cell products

### Second generation CARs



### Co-stimulation

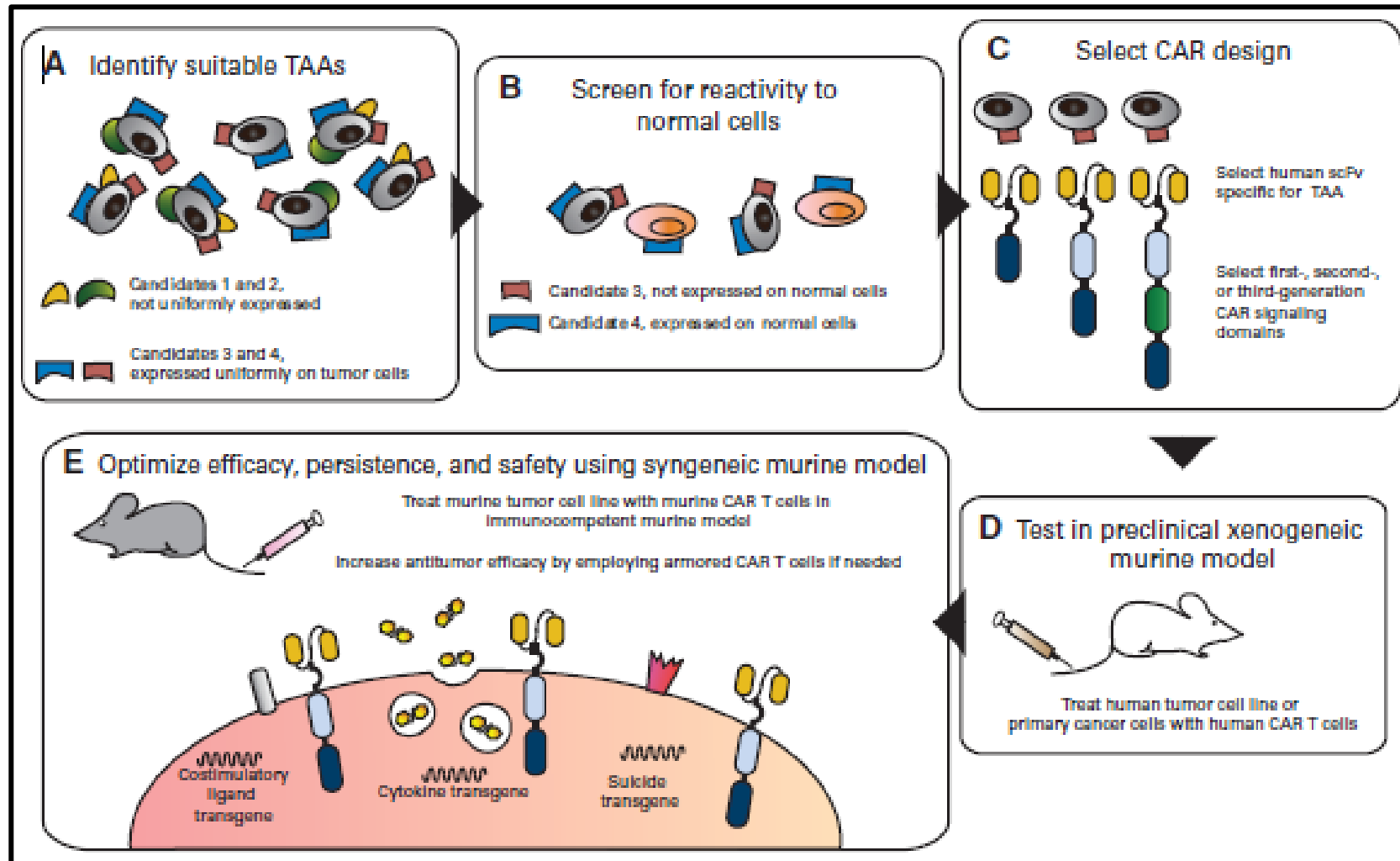
- CD28  
faster in vivo expansion
- higher peak levels
- 4-1BB  
drive towards CM phenotype
- longer persistence
- less exhaustion





# CAR-T cells

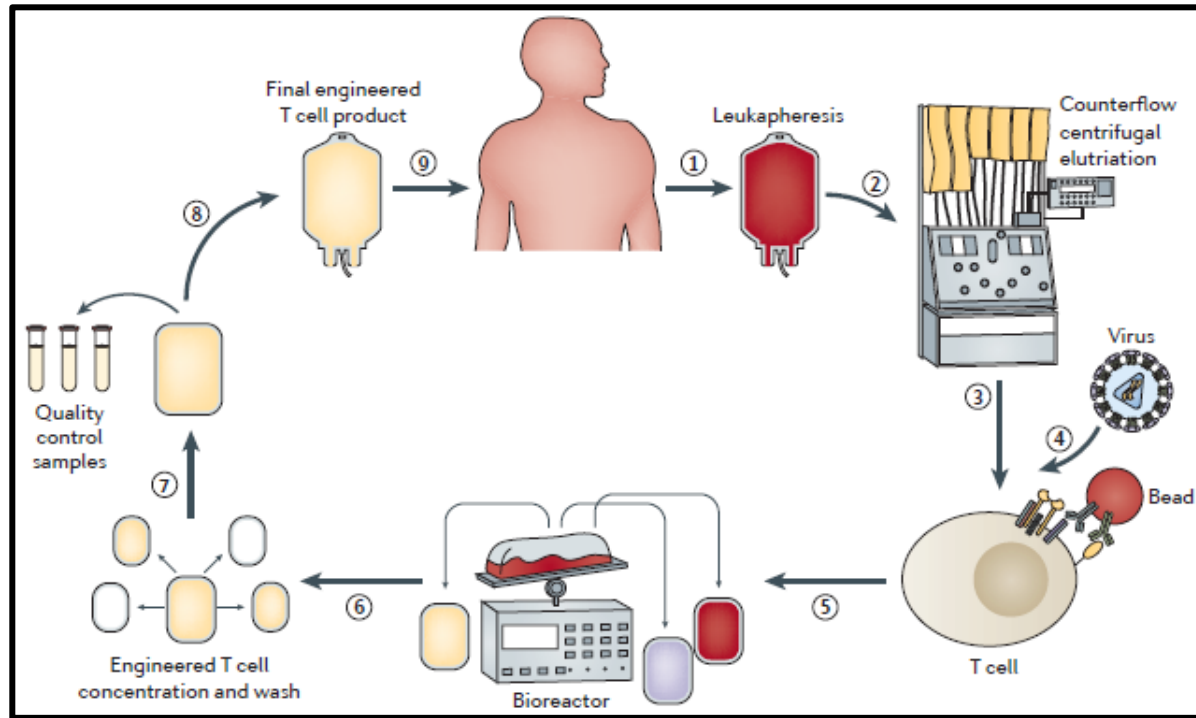
## Academic production : pre-clinical development





# CAR-T cells

## Academic production : CAR-T manufacturing



**Table 8.3** CAR-T manufacturing methodology

	Potential methods	Timepoint
Step 1: T cell enrichment post-leukapheresis ( <i>optional</i> )	Ficoll density gradient centrifugation; elutriation; immunomagnetic bead separation	Day 1
Step 2: T cell activation using synthetic antigen presenting technologies (CD3 +/- CD28) ( <i>required</i> )	Soluble monoclonal antibodies; Para-magnetic anti-CD3/CD28 antibody coated beads; polymeric biodegradable CD3/28 incorporating nanomatrix (TransAct™)	Days 1, 2
Step 3: T cell stimulation ( <i>required</i> )	IL-2, IL-7, and IL-15 in the culture medium (as per protocol) (Hoffmann et al. 2018; Gong et al. 2019)	From day 1 onwards
Step 4: Gene delivery/transduction with a retroviral or lentiviral CAR vector ( <i>required</i> )	In some processes, retronectin or Vectofusin® is used to enhance transduction ( <i>optional</i> )	Days 2, 3
Step 5: T cell expansion ( <i>required</i> )	T-flasks, plates or culture bags; bioreactors, e.g., G-Rex™ flask (Wilson Wolf Manufacturing); Xuri WAVE™ Bioreactor (GE Life Systems); CliniMACS Prodigy™ (Miltenyi BioTec)	Days 3, 4 and onwards
Step 6: T cell harvest and cryopreservation ( <i>required</i> )	The cryopreservation methodology often mirrors processes defined for haematopoietic cells. Methods include passive freezing (-80 °C freezer) and controlled-rate freezing	Day 8 onwards
Step 7: CAR-T cell quality assurance control and release testing	In-process and end of process controls are taken to ensure the product complies with release criteria specifications	Day 8 onwards

# CAR-T cells

## Academic production : QC

### Vector production

**Table 8.1** Quality control for the HEK293T master cell bank

Parameter	Method	Acceptance criteria
Appearance	Visual inspection	Presence of adherent cells with thin extensions
Sterility	Microbial growth	Sterile
Mycoplasma	PCR	Absent
Adventitious viruses	PCR	Absent
Karyotype	G-band staining	Informative
Cell viability (%) after thawing	Neubauer cell counting with trypan blue exclusion	>70%

**Table 8.2** Quality control for GMP-grade virus production

Parameter	Method	Acceptance criteria
Appearance	Visual inspection	Yellowish liquid solution
Viral titre	Limiting dilution	$>3.75 \times 10^7$ TU/mL
Sterility	Microbial growth	Sterile
Mycoplasma	PCR	Absent
Identity	PCR	Positive
Replication-competent lentivirus	Real-time PCR	Absent

### CAR-T cell production

**Table 8.4** Quality control of CAR-T cell biology and microbiology

Parameter	Method	Acceptance criteria
Appearance	Visual inspection	Cloudy liquid solution
CAR+ cells (%) <sup>a</sup>	Flow cytometry	>20%
CD3+ cells (%)	Flow cytometry	>70%
Cell viability (%)	Neubauer cell counting with trypan blue exclusion <sup>b</sup>	>70%
Sterility	Microbial growth E. Ph. 2.6.1	Sterile from bacteria/fungi
Mycoplasma	PCR <sup>c</sup>	Absent
Endotoxin	Chromogenic assay	<0.5 EU/mL
<i>Optional/R&amp;D</i>		
CAR/CD45RA/CCR7 For detection of TE/ TEM/TEMRA/TCM/TN subpopulations	Flow cytometry	A high proportion of immature T cells is desirable for a long-lasting CAR-T cell effect in the patient
Cytotoxic potency	Cr-51 release assays in tumour CAR-T cell co-culture, assessed by flow cytometry	>40% killing at an effector/target ratio of 10:1 (or higher ratio) in a 4-h assay
Adventitious viruses	PCR	Absent
Number of transgene copies/cell	Real-time PCR (Kunz et al. 2019; Schubert et al. 2020)	<10 (range <7–15!) copies/cell <sup>d</sup>

<sup>a</sup>Automated cell counters, such as Luna™, are highly recommended

<sup>b</sup>Highly specific detection reagents (e.g., the Miltenyi Detection Reagent™) are strongly advised to distinguish CAR-T cells from the negative fraction

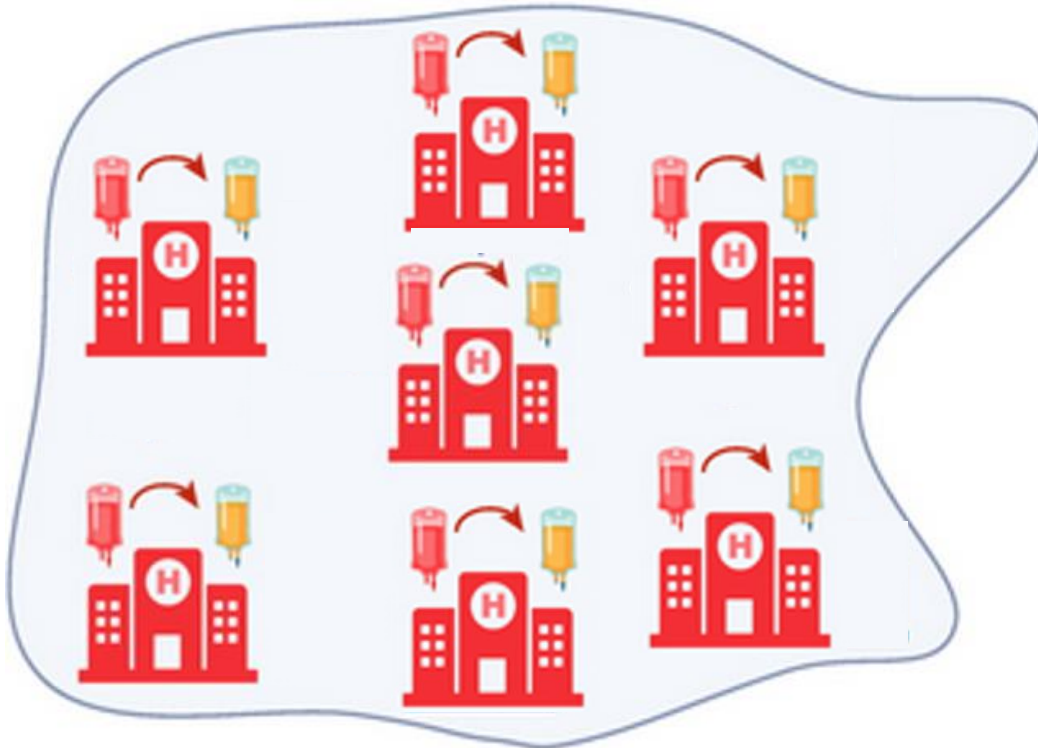
<sup>c</sup>European standards stipulate PCR methodology, in contrast to US regulations, which require serology

<sup>d</sup>Differs between countries and products

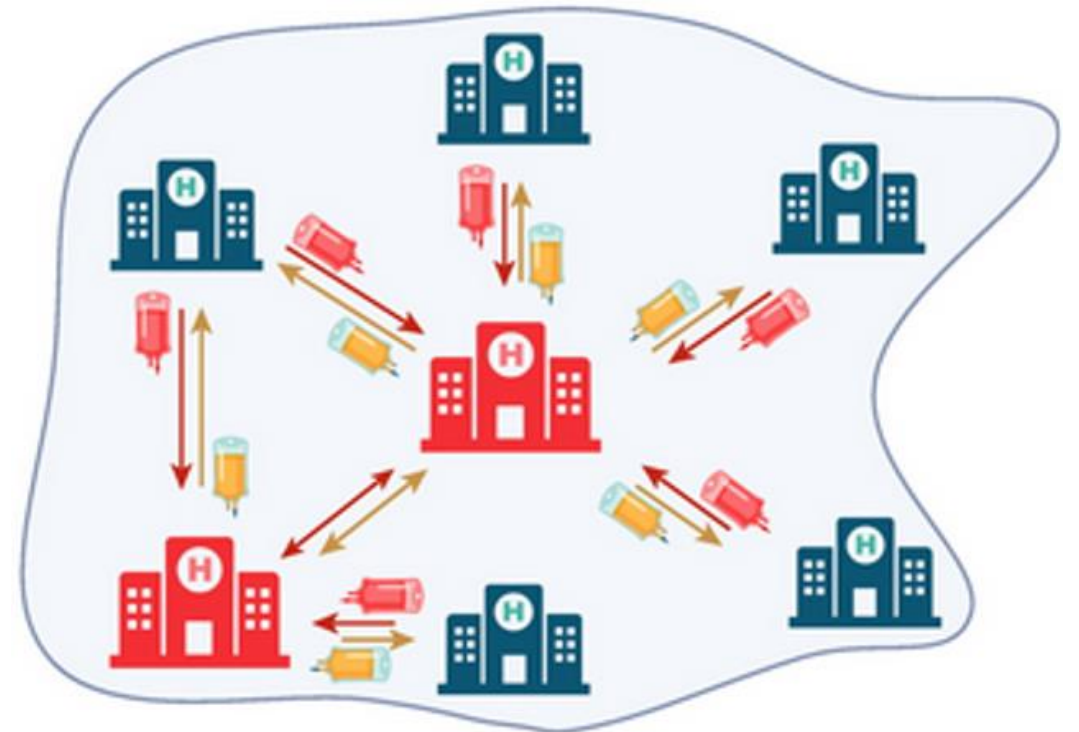
# CAR-T cells

## Point-of-care & decentralized CAR-T cell production

Point-of-care



Decentralized



# CAR-T cells

## Galapagos decentralized production

*Enabling scalable and consistent decentralized production*



### Clinical trials

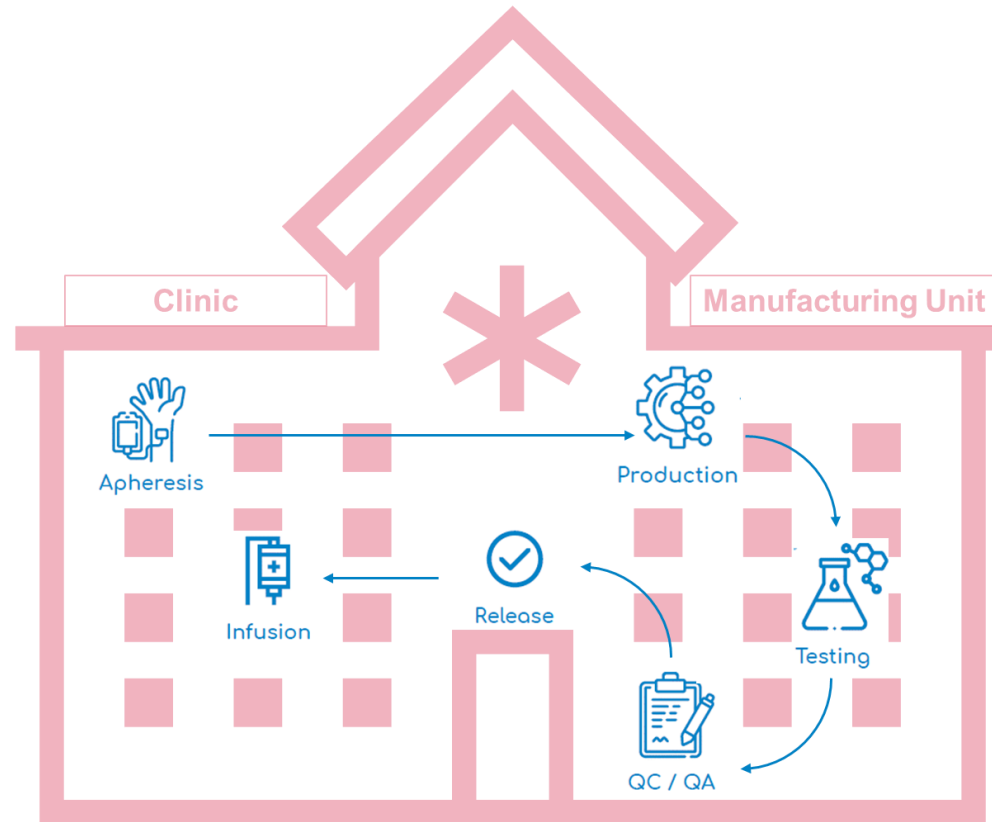
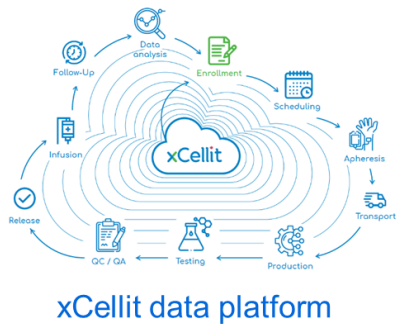
- NHL : Atalanta-1
- CLL / RT : Euplagia-1
- MM : Papilio-1

# CAR-T cells

## Galapagos decentralized production

### Manufacturer

- Study Sponsor
- Real-time data capture
- Product responsibility

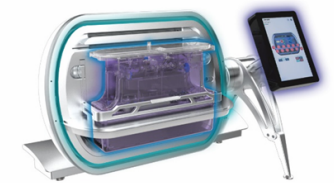


### Site selection:

- Clinical CAR-T experience
- Cleanroom facility (class B-C)
- Operators

### Tech transfer & support:

- Cocoon instruments
- QC instruments
- Materials kits
- Procedures & training
- Service & support



Cocoon

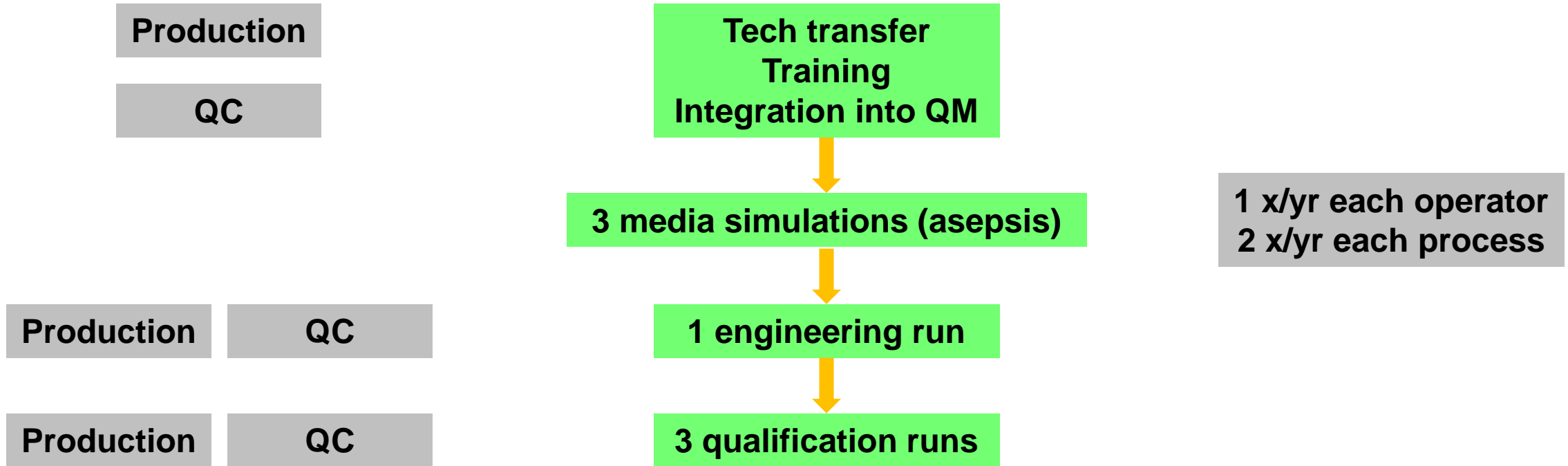


Rapid QC instruments



# CAR-T cells

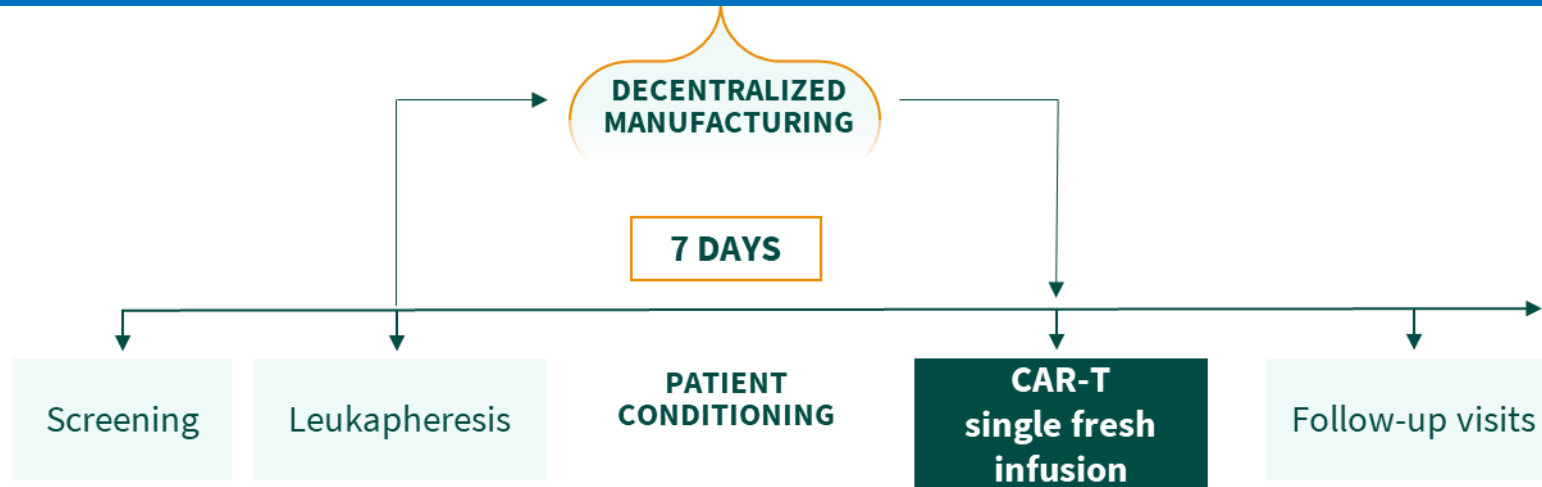
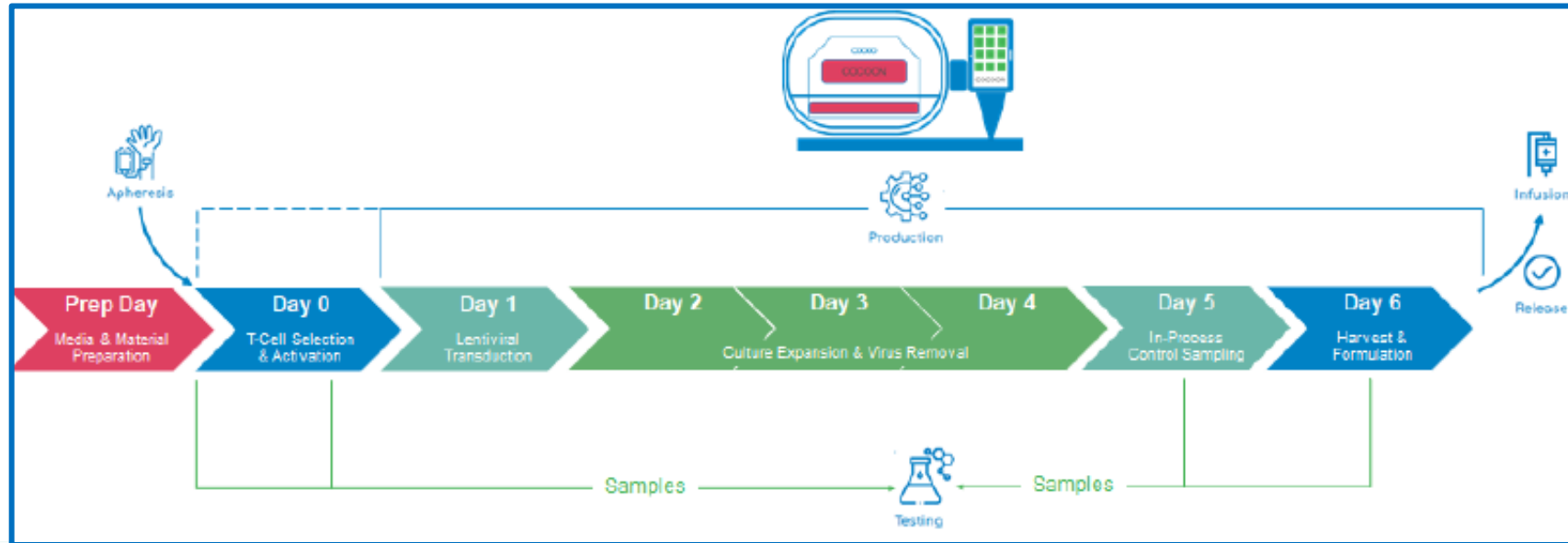
## Production qualification for phase I-II



**Process qualification if all within specifications !**

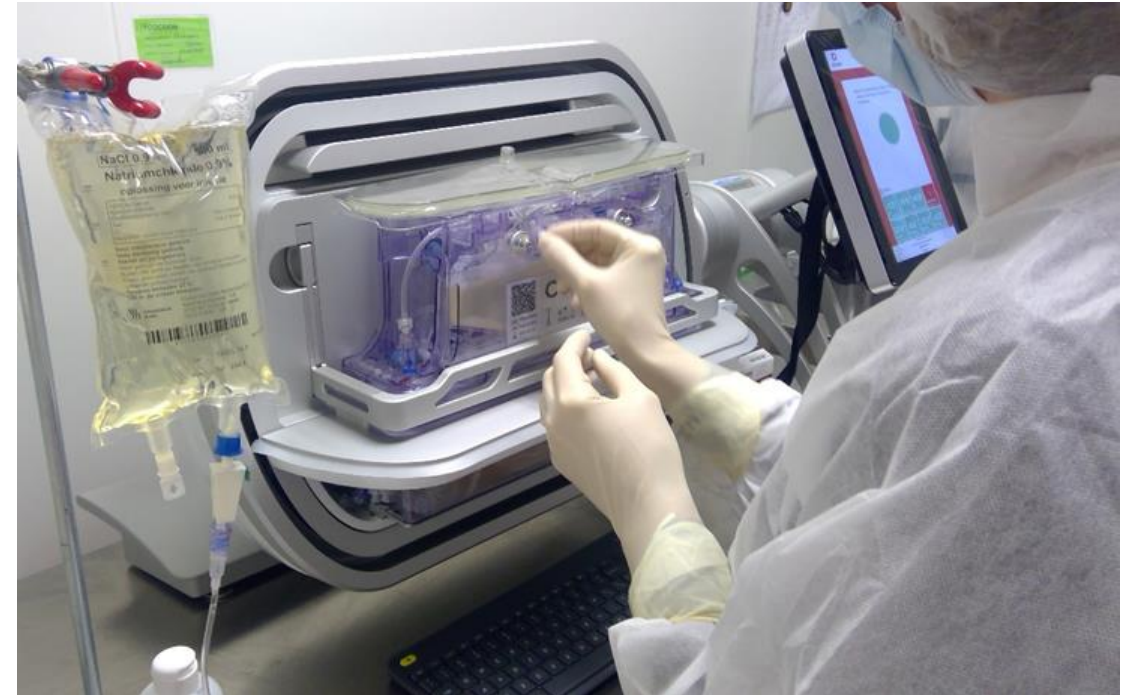
# CAR-T cells

## Galapagos decentralized production & patient journey



# CAR-T CELLS

## Production on Cocoon automated platform



# **CAR-T CELL THERAPY**

**CLINICAL TRIALS**

**NHL : ATALANTA**

# Seven-Day Vein-to-Vein Point-of-Care Manufactured CD19 CAR T Cells (GLPG5101) in R/R NHL: Results from the Phase 1/2 ATALANTA-1 Trial

MJ Kersten *et al*

Amsterdam, Leiden, Antwerp, Liège

Galapagos

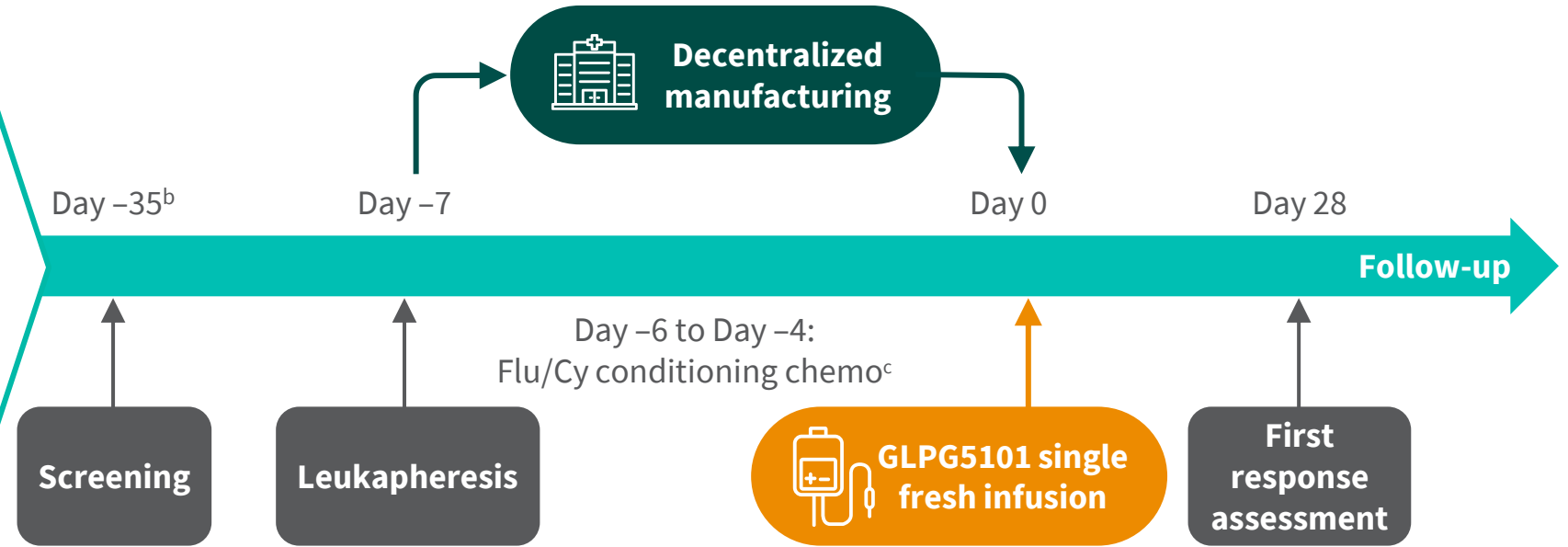
**EHA 2024**



# ATALANTA-1 Study Design and Objectives

## Key eligibility criteria

- No prior CD19-targeted therapies
- Phase 1 dose escalation:**
- DLBCL
    - Primary refractory or first relapse
  - FL, MZL, MCL
    - Relapsed or refractory after two prior treatments
- Phase 2 expansion cohorts:**
- DLBCL, HR DLBCL,<sup>a</sup> FL + MZL, MCL, Burkitt lymphoma, PCNSL



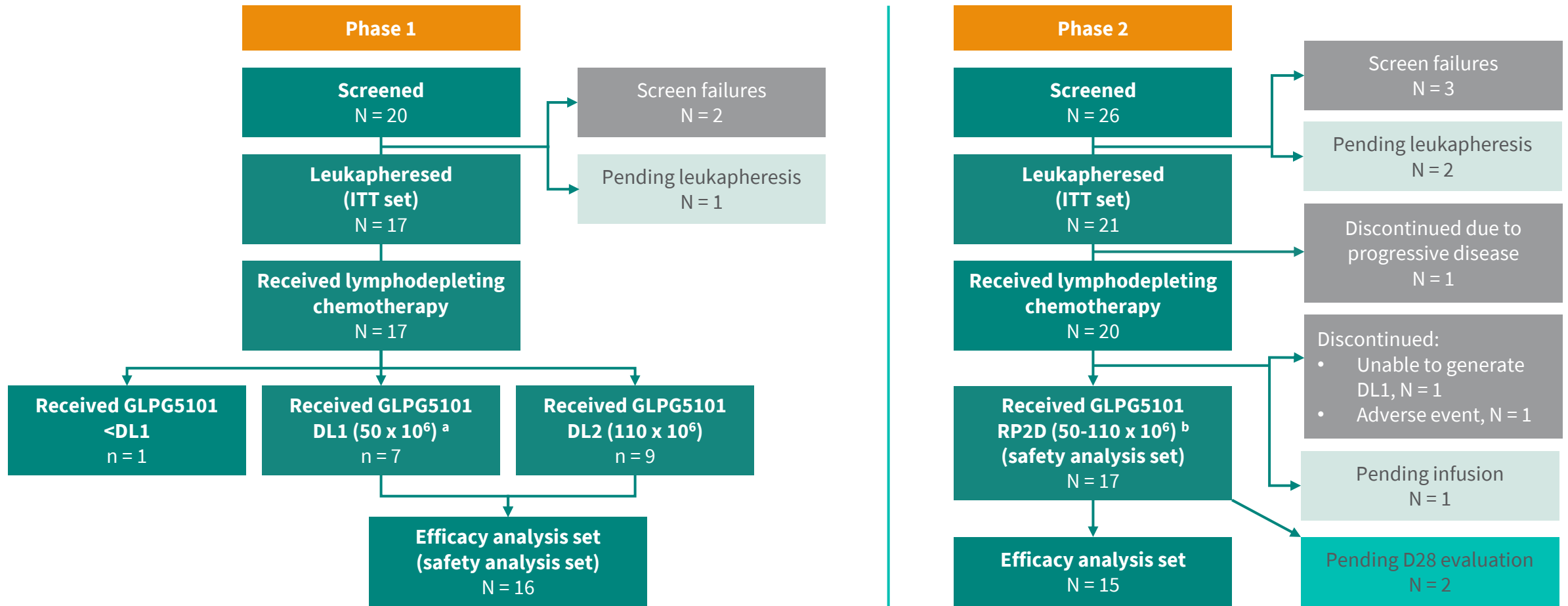
**Phase 1 primary objectives:**  
Safety  
Determination of a RP2D

**Phase 2 primary objective:**  
Efficacy (ORR)

**Phase 1/2 secondary objectives:**  
Safety  
Efficacy (CRR, DoR, MRD-, PFS, OS)  
Pharmacokinetics and pharmacodynamics  
Feasibility of decentralized manufacturing

<sup>a</sup>IPI 3-5 or double/triple-hit lymphoma. <sup>b</sup>Screening could take place up to a maximum of 28 days prior to leukapheresis. <sup>c</sup>Conditioning chemotherapy: fludarabine IV (30 mg/m<sup>2</sup>/day); cyclophosphamide IV (300 mg/m<sup>2</sup>/day).  
Cy, cyclophosphamide; FL, follicular lymphoma; Flu, fludarabine; (HR) DLBCL, (high-risk) diffuse large B-cell lymphoma; IPI, international prognostic index; IV, intravenous; MCL, mantle cell lymphoma;  
MZL, marginal zone lymphoma; ORR, objective response rate; PCNSL, primary central nervous system lymphoma; RP2D, recommended Phase 2 dose

# Patient Disposition



<sup>a</sup>Includes 3 patients who received DL1 instead of planned DL2, due to lower manufacturing yield.

<sup>b</sup>Sixteen patients received RP2D; one patient received <RP2D.

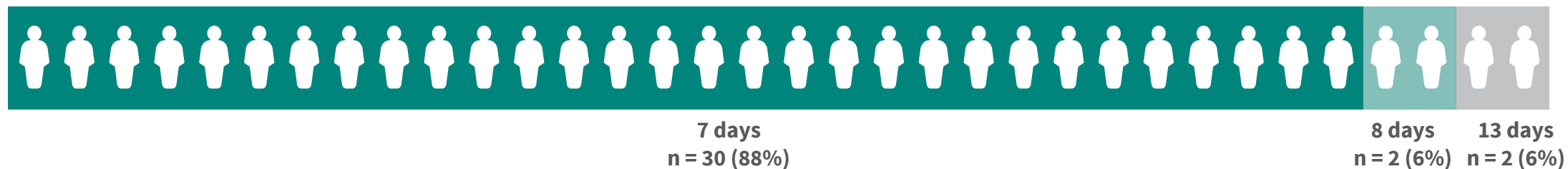
DL, dose level; ITT, intention-to-treat; RP2D, recommended Phase 2 dose

Data cutoff: December 20, 2023.

# Decentralized Manufacturing

*Enabling fresh product infusion with a 7-day vein-to-vein time*

Median vein-to-vein time for product was **7 days** (range 7–13)



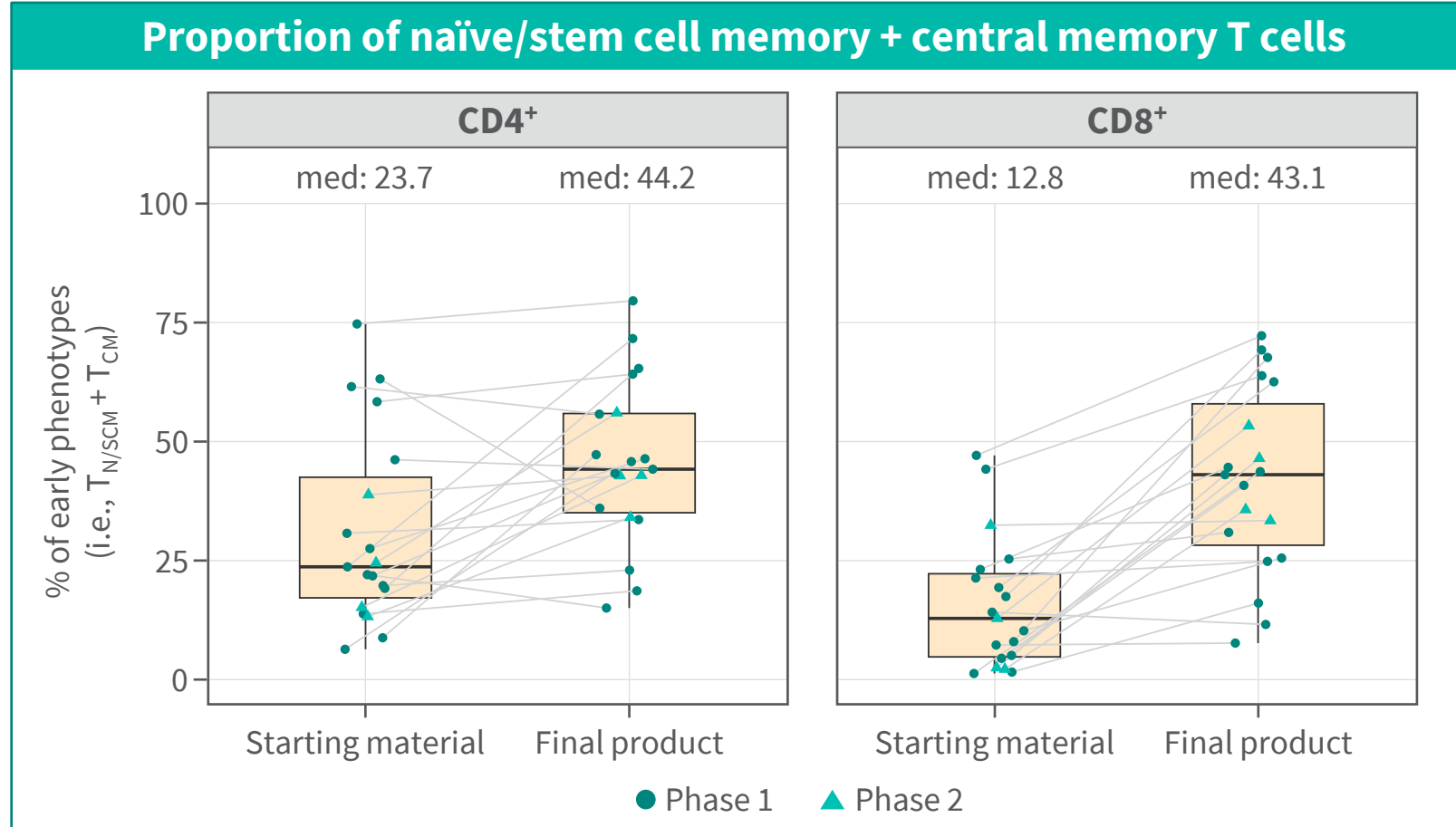
- Short vein-to-vein time eliminated the need for bridging therapy
- GLPG5101 was administered as a **fresh product** to 32/34 (94%) patients
  - Two patients received a cryopreserved product (vein-to-vein time 13 days)

Data cutoff: December 20, 2023.

Adapted from presentation at EHA Hybrid Congress: 13–16 June, 2024; Madrid, Spain

# Product Characterization

*Proportion of early phenotypes of CD4<sup>+</sup> and CD8<sup>+</sup> CAR T cells was increased in the final product compared with the starting material*



Early phenotypes of  
CD4<sup>+</sup> & CD8<sup>+</sup> CAR T cells

=

Naïve/stem cell memory T cells  
(CD45RO<sup>-</sup>CD197<sup>+</sup> T<sub>N/SCM</sub>)

+

Central memory T cells  
(CD45RO<sup>+</sup>CD197<sup>+</sup> T<sub>CM</sub>)

Exploratory flow cytometry analysis of T-cell subsets in the apheresis starting material and final product (gated on CAR<sup>+</sup> T cells for final product) for paired patient samples (N=19).  
CAR, chimeric antigen receptor; med, median; T<sub>CM</sub>, central memory T cells; T<sub>N/SCM</sub>, naïve/stem cell memory T cells

Data cutoff: September 01, 2023.

# Demographics and Baseline Characteristics

*High-risk and heavily pretreated patients were included*

	Phase 1 (DL1 & 2) All patients N = 16	Phase 2 All patients N = 17
Age, median (range), years	65 (25-77)	67 (45-81)
Male, n (%)	12 (75)	9 (53)
NHL subtype, n (%) <sup>a</sup>		
DLBCL	9 (56)	0
MCL <sup>b</sup>	3 (19)	4 (24)
FL	3 (19)	12 (71)
MZL	1 (6)	1 (6)
IPI/MIPI/FLIPI score at screening, high risk, n (%)	6 (38)	11 (65)
ECOG PS at baseline, n (%)		
0	6 (38)	8 (47)
1	9 (56)	6 (35)
2	1 (6)	3 (18)
Previous therapies, median (range)	3 (1-7)	3 (2-11)
Ann Arbor disease stage, n (%)		
II	1 (6)	4 (24)
III-IV	15 (94)	13 (76)

<sup>a</sup>Sum of percentages may be >100 due to rounding. <sup>b</sup>Two patients with MCL were not included in the Phase 2 efficacy analysis set as the first response assessment data were not available at data cutoff.

Data cutoff: December 20, 2023.

DLBCL, diffuse large B-cell lymphoma; ECOG PS, Eastern Cooperative Oncology Group performance status; FL, follicular lymphoma; (M, FL)IPI, (MCL, FL) international prognostic index; MCL, mantle cell lymphoma; MZL, marginal zone lymphoma; NHL, non-Hodgkin lymphoma

Adapted from presentation at EHA Hybrid Congress: 13-16 June, 2024; Madrid, Spain



# Safety: TEAEs

*Most Grade  $\geq 3$  TEAEs were hematological*

TEAEs up to 14 weeks after infusion	Phase 1 (DL1 & 2)	Phase 2
	All patients N = 16	All patients N = 17
Any TEAE, n (%)	16 (100)	17 (100)
Any GLPG5101-related TEAE, n (%)	16 (100)	14 (82)
Serious TEAE, n (%)	5 (31)	3 (18)
TEAE leading to death, n (%)	1 (6)	0
Any Grade $\geq 3$ TEAE, n (%)	16 (100)	14 (82)
<b>Hematological Grade <math>\geq 3</math> TEAEs, n (%)</b>		
Neutropenia <sup>a</sup>	15 (94)	12 (71)
Anemia <sup>b</sup>	6 (38)	1 (6)
Lymphopenia <sup>c</sup>	5 (31)	3 (18)
Thrombocytopenia <sup>d</sup>	4 (25)	4 (24)
Leukopenia <sup>e</sup>	6 (38)	5 (29)
<b>Other Grade <math>\geq 3</math> TEAEs in <math>\geq 2</math> patients<sup>f</sup>, n (%)</b>		
Pyrexia	2 (13)	1 (6)
Pleural effusion	2 (13)	0

<sup>a</sup>Includes neutropenia/neutrophil count decreased. <sup>b</sup>Includes anemia/hemoglobin decreased. <sup>c</sup>Includes lymphopenia/lymphocyte count decreased. <sup>d</sup>Includes thrombocytopenia/platelet count decreased. <sup>e</sup>Includes leukopenia/white blood cell count decreased. <sup>f</sup>In either the Phase 1 or Phase 2 total population.  
 TEAE, treatment-emergent adverse event

**Data cutoff: December 20, 2023.**

# Safety: AESIs and Deaths

*The vast majority of CRS and ICANS events were low-grade*

AESIs up to 14 weeks after infusion	Phase 1 (DL1 & 2)	Phase 2
	All patients N = 16	All patients N = 17
<b>CRS (n, %)</b>	<b>7 (44)</b>	<b>5 (29)</b>
Grade 1	2 (13)	4 (24)
Grade 2	3 (19)	1 (6)
Grade 3	2 (13)	0
<b>ICANS (n, %)</b>	<b>6 (38)</b>	<b>1 (6)</b>
Grade 1	6 (38)	0
Grade 2	0	0
Grade 3	0	1 (6)
<b>Infections, Grade ≥3 (n, %)</b>	<b>1 (6)</b>	<b>0</b>
<b>Prolonged cytopenia, <sup>a</sup> Grade ≥3, (n,%)</b>		
30 days after infusion <sup>b</sup>	7 (47)	5 (36)
60 days after infusion <sup>c</sup>	4 (27)	3 (27)
<b>Hemophagocytic lymphohistiocytosis, any grade (n, %)</b>	<b>1 (6)</b>	<b>0</b>

**CRS and ICANS**

Two cases of Grade 3 CRS in Phase 1

One case of Grade 3 ICANS in Phase 2

**Deaths during treatment (up to 14 wks after infusion)**

Intra-abdominal hemorrhage, caused by DIC  
Phase 1, DL2<sup>e</sup>

Respiratory distress, caused by disease progression & respiratory infection  
Phase 1, <DL1<sup>d</sup>

**Deaths post-treatment period<sup>g</sup>:**

Escherichia sepsis  
Phase 1, DL2<sup>e,f</sup>

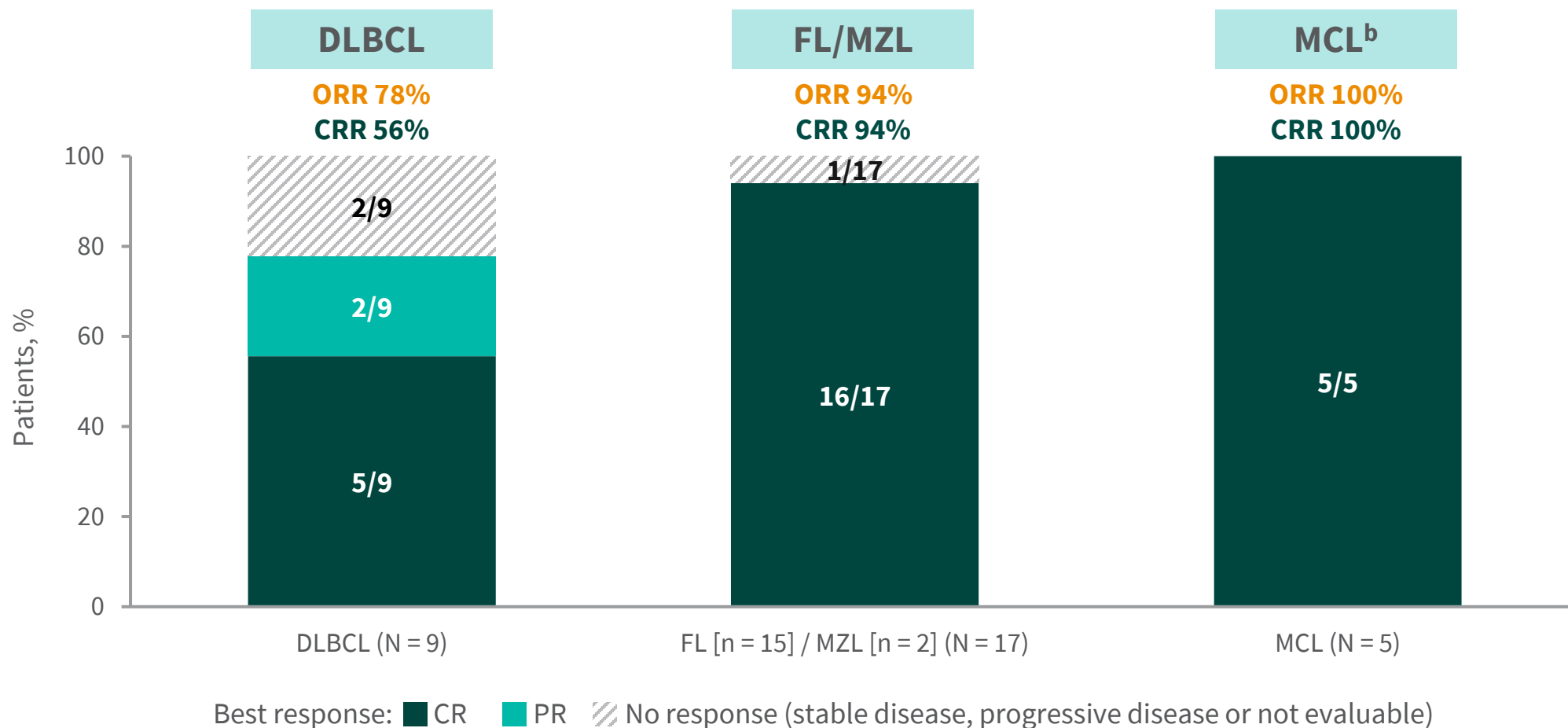
Data cutoff: December 20, 2023.

<sup>a</sup>Includes all events related to neutropenia, thrombocytopenia, anemia and lymphopenia. <sup>b</sup>Data available for 15 patients in Phase 1 and 14 patients in Phase 2. <sup>c</sup>Data available for 15 patients in Phase 1 and 11 patients in Phase 2.

<sup>d</sup>DL1 = 50×10<sup>6</sup> CAR+ T cells. <sup>e</sup>DL2 = 110×10<sup>6</sup> CAR+ T cells. <sup>f</sup>Reported >6 months post-infusion, in a patient with hypogammaglobulinemia. AESI, adverse event of special interest; CRS, cytokine release syndrome; DIC, disseminated intravascular coagulation; DL, dose level; ICANS, immune effector cell-associated neurotoxicity syndrome

# Efficacy: Pooled Phase 1/2 Results

*High OR and CR rates were observed<sup>a</sup>*



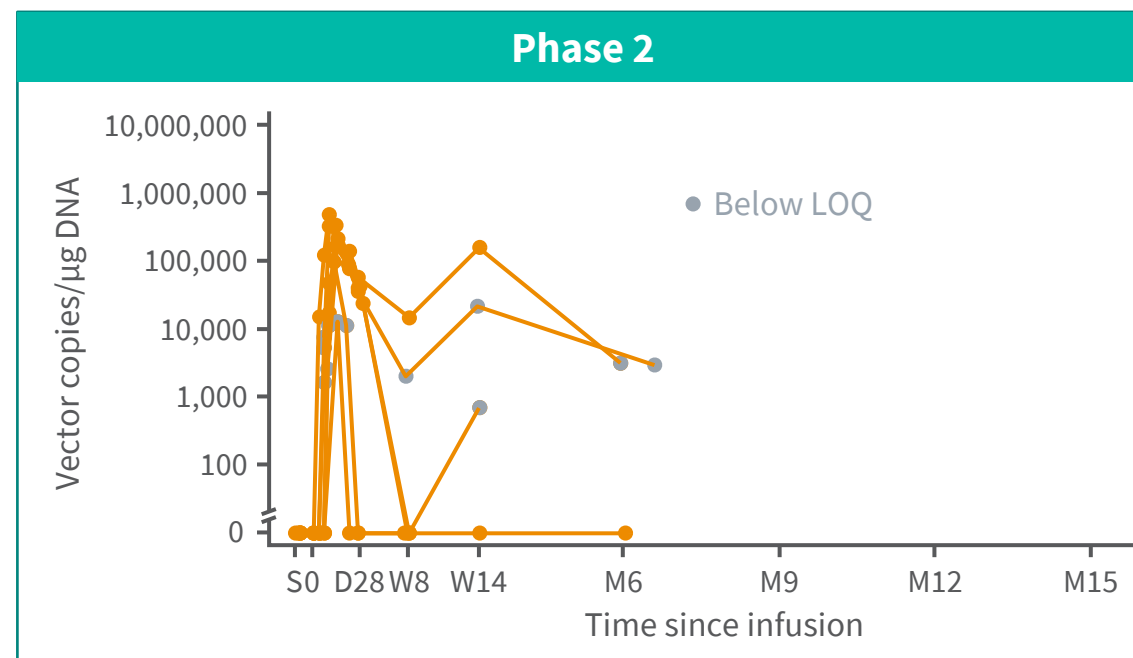
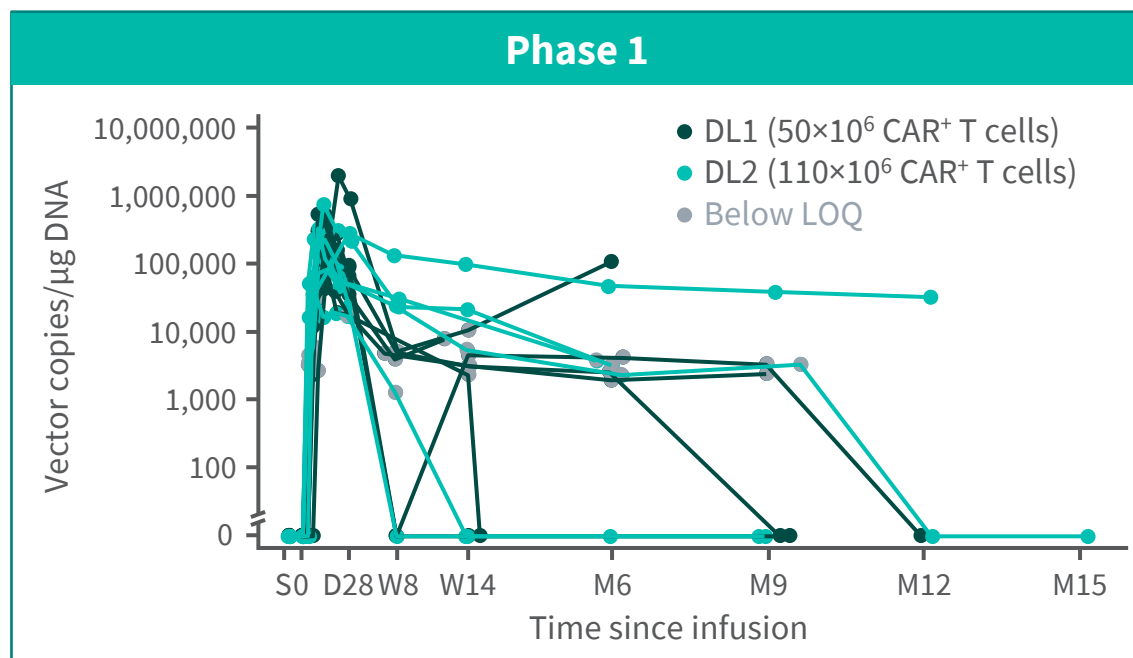
<sup>a</sup>Best response at any time after infusion. <sup>b</sup>Two patients with MCL were not included in the Phase 2 efficacy analysis set as the first response assessment data were not available at data cutoff. **Data cutoff: December 20, 2023.**

CR, complete response; CRR, complete response rate; DLBCL, diffuse large B-cell lymphoma; FL, follicular lymphoma; MCL, mantle cell lymphoma; MZL, marginal zone lymphoma; OR, objective response; ORR, objective response rate; PR, partial response



# Cellular Kinetics

*GLPG5101 demonstrated the ability for durable persistence and robust expansion across doses*



**11/15 (73%) patients had detectable GLPG5101 in peripheral blood at Week 14 post-infusion**

Persisting CAR T cells could be detected up to 12 months post-infusion

Quantification of GLPG5101 in peripheral blood by qPCR. LOQ: 1,000 vector copies.

Data cutoff: December 20, 2023.

CAR, chimeric antigen receptor; D, Day; DL, dose level; LOQ, limit of quantification; M, Month; S, screening; W, Week

Adapted from presentation at EHA Hybrid Congress: 13–16 June, 2024; Madrid, Spain

# Conclusions and Study Updates

- Data from **33 patients with relapsed/refractory NHL** enrolled in the ongoing Phase 1/2 ATALANTA-1 study demonstrate that **decentralized CAR T-cell manufacturing with a short vein-to-vein time is feasible**
- GLPG5101 was administered as a **fresh and fit product** with a **median vein-to-vein time of 7 days**
- GLPG5101 demonstrated **robust *in vivo* expansion and durable persistence** post-infusion
- The **vast majority of CRS and ICANS events were Grade 1 or 2; two cases of Grade 3 CRS and one case of Grade 3 ICANS** were reported
- **High complete response rates were observed across indications** in this heavily pretreated population

## Study updates

- The RP2D in FL, MZL and MCL is DL2 (110 (range 50-110) x 10<sup>6</sup> CAR+ T cells)
- Dose escalation in DLBCL is ongoing at DL3 (250 x 10<sup>6</sup> CAR+ T cells)
- Additional expansion cohorts of patients with BL<sup>a</sup> and PCNSL<sup>a</sup> will be treated at the RP2D for DLBCL

<sup>a</sup>After receiving two or more prior treatments.

BL, Burkitt lymphoma; DL, dose level; DLBCL, diffuse large B-cell lymphoma; FL, follicular lymphoma; MCL, mantle cell lymphoma; MZL, marginal zone lymphoma; PCNSL, primary central nervous system lymphoma RP2D, recommended Phase 2 dose; CAR, chimeric antigen receptor; CRS, cytokine release syndrome; ICANS, immune effector cell-associated neurotoxicity syndrome; NHL, non-Hodgkin lymphoma

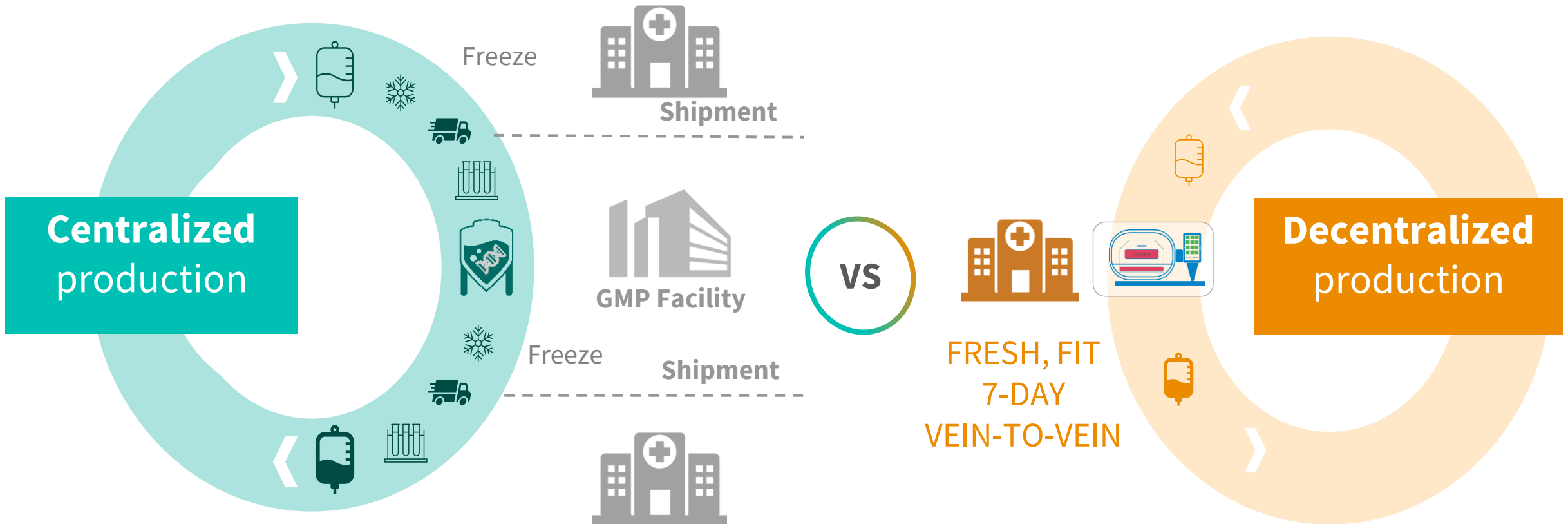


# **CAR-T CELL THERAPY**

## **DISCUSSION**

# CAR-T cells

## Commercial vs decentralized production



# CAR-T cells

## Decentralized production : advantages

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- **Potential advantages**

- Fresh product
- Short vein-to-vein time (7 days)
- No need for bridging therapy
- Simplified logistics
- Potentially lower costs
- Improved patient access
  
- Close partnership between industry & hospital
- Very motivating for Lab of Cell & Gene Therapy
- Integrated production/QC & clinical use



# CAR-T cells

## Decentralized production : difficulties

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- **Potential difficulties**

- Requirement for academic GMP manufacturing & QC site : facilities, equipment, reagents, staff, QM system, accreditation by regulatory authorities
- If ATMP comes from academic research :
  - Pre-clinical product development
  - Validated manufacturing & QC processes
  - Marketing authorization & pharmacovigilance
  - Funding
- If ATMP comes from biotech company :
  - Tech transfer, training & validation at each site (production & QC)
  - Standardization across multiple sites (production & QC)
  - Quality agreement to define respective responsibilities
  - Financial agreement
- Clinical trials (phase 1-2 vs phase 3) versus commercialization

# CAR-T cells

## Decentralized production : responsibilities

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- **Research : clinical trials → sponsor**
- **Commercialization → biotech company vs production site ?**

### Items

- **Management of reagents (other than viral vector) & equipment**
  - **Manufacturing issues : out-of-specification, failure, change control**
  - **QC issues : decentralized vs centralized, change control**
  - **Patient issues : drop-out, unexpected AE, long-term pharmacovigilance**
- **Legal & financial responsibility**

# CAR-T cells

## Decentralized production : cost structure

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- **Biotech company = MA holder**
  - IP on reagents : viral vector...
  - IP on whole production & QC processes
  - Centralized data platform
  - Long-term pharmacovigilance & unexpected AE
- **Production site**
  - Facility building & maintenance
  - Staff recruitment & training
- **To be decided / shared**
  - Equipment purchase & maintenance
  - Management of reagents (other than the ones covered by Biotech IP)
  - Responsibilities for OOS, manufacturing failures, patient drop-out



**THANK YOU**

**FOR YOUR ATTENTION !**